

TOPIC 12

CONSTRAINED ORDINATION



Constrained ordinations

- constrained ordination adds a level of statistical testing (next topic)
- also called **direct gradient analysis** or **canonical ordination**
- uses an **explanatory matrix** to explain the patterns (or variability) in the **response matrix**
- (A) ...two matrices; explanatory grouping variable(s) (**env.; qualitative, ordinal**), and a multivariate response (**spp.**)
 - Discriminant Analysis (**DA**)
 - **✗** Analysis of Similarities (**ANOSIM**)
 - **✓** Permutational Analysis of Variance (**PERMANOVA**)
 - Mantel test
 - etc...
- (B) ...two matrices; a multivariate explanatory variables (**env.; continuous, categorical, or ordinal covariate(s)**), and a multivariate response (**spp.**)
 - Redundancy Discriminant Analysis (**RDA**) – ‘extension’ of PCA
 - **✓** Distance-based redundancy analysis (**db-RDA**)
 - **✓** Canonical Correspondence Analysis (**CCA**) – extension of CA
 - Canonical Analysis of Principal Coordinates (**CAP**) – extension of PCoA

Constrained ordinations

- explanatory variables sometimes called external variables
- used in an **hypothesis-driven** setting
- CCA and db-RDA combine classical ordination (CA and PCA, respectively) with multiple regression
- the significance of the ordinations is assessed via permutation (bootstrapping) tests, e.g. as in a PERMANOVA

Multivariate Techniques

Obs	Group	X-set					Y-set				
1	A	a_{11}	a_{12}	a_{13}	...	a_{1p}	b_{11}	b_{12}	b_{13}	...	b_{1m}
2	A	a_{21}	a_{22}	a_{23}	...	a_{2p}	b_{21}	b_{22}	b_{23}	...	b_{2m}
3	A	a_{31}	a_{32}	a_{33}	...	a_{3p}	b_{31}	b_{32}	b_{33}	...	b_{3m}
.	
.	
n	A	a_{n1}	a_{n2}	a_{n3}	...	a_{np}	b_{n1}	b_{n2}	b_{n3}	...	b_{nm}
n+1	C	c_{11}	c_{12}	c_{13}	...	c_{1p}					
n+2	C	c_{21}	c_{22}	c_{23}	...	c_{2p}					
n+3	C	c_{31}	c_{32}	c_{33}	...	c_{3p}					
.					
.					
N	C	c_{n1}	c_{n2}	c_{n3}	...	c_{np}					


factors

(A) Constrained ordination
 Discriminant Analysis (DA)
 Analysis of Similarities (ANOSIM)
 Permutational ANOVA (PERMANOVA)
 Mantel test
 etc...

Multivariate Techniques

Obs	Group	X-set				Y-set			
1	A	a_{11}	a_{12}	a_{13}	... a_{1p}	b_{11}	b_{12}	b_{13}	... b_{1m}
2	A	a_{21}	a_{22}	a_{23}	... a_{2p}	b_{21}	b_{22}	b_{23}	... b_{2m}
3	A	a_{31}	a_{32}	a_{33}	... a_{3p}	b_{31}	b_{32}	b_{33}	... b_{3m}
·	·	·	·	·	... ·	·	·	·	... ·
·	·	·	·	·	... ·	·	·	·	... ·
n	A	a_{n1}	a_{n2}	a_{n3}	... a_{np}	b_{n1}	b_{n2}	b_{n3}	... b_{nm}
n+1	C	c_{11}	c_{12}	c_{13}	... c_{1p}				
n+2	C	c_{21}	c_{22}	c_{23}	... c_{2p}				
n+3	C	c_{31}	c_{32}	c_{33}	... c_{3p}				
·	·	·	·	·	... ·				
·	·	·	·	·	... ·				
N	C	c_{n1}	c_{n2}	c_{n3}	... c_{np}				

continuous



(B) Constrained ordination
 Redundancy Discriminant Analysis (RDA)
 Canonical Correspondence Analysis (CCA)
 Distance-based redundancy analysis (db-RDA)
 Canonical analysis of principal coordinates (CAP)

Redundancy Analysis (RDA)

- see <https://sites.google.com/site/mb3gustame/constrained-analyses/rda> and https://www.davidzeleny.net/anadat-r/doku.php/en:rda_cca
- a method that combines PCA with multiple regression
- it is therefore a 'linear' method, and the same constraints inherent in a PCA are present here too
- in essence, it is a multivariate (multiple response) multiple linear regression, followed by a PCA of the table of fitted values
- it works on centered response data (species, matrix **Y**) and standardised explanatory data (env., etc., matrix **X**)
- **X** conditions the weights (eigenvalues), the orthogonality, and the direction of the ordination axes
- an extremely useful and powerful method available to ecologists
- additional detail in 6.3.1. of Numerical Ecology with R

```
> spe
# A tibble: 29 x 27
  Cogo Satr Phph Babl Thth Teso Chna Pato Lele Sqce Baba Albi Gogo Eslu Pefl Rham Legi Scer Cyca Titi Abbr Icme Gyce
<int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int> <int>
1     0     3     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0
2     0     5     4     3     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0
3     0     5     5     5     0     0     0     0     0     0     0     0     0     1     0     0     0     0     0     0     0     0     0     0
4     0     4     5     5     0     0     0     0     0     0     1     0     0     1     2     2     0     0     0     0     1     0     0     0
5     0     2     3     2     0     0     0     0     0     5     2     0     0     2     4     4     0     0     2     0     3     0     0     0
6     0     3     4     5     0     0     0     0     0     1     2     0     0     1     1     1     0     0     0     0     2     0     0     0
7     0     5     4     5     0     0     0     0     0     1     1     0     0     0     0     0     0     0     0     0     0     0     0     0
8     0     0     1     3     0     0     0     0     0     0     5     0     0     0     0     0     0     0     0     0     1     0     0     0
9     0     1     4     4     0     0     0     0     0     2     2     0     0     1     0     0     0     0     0     0     0     0     0     0
10    1     3     4     1     1     0     0     0     0     0     1     0     0     0     0     0     0     0     0     0     0     0     0     0
# ... with 19 more rows, and 4 more variables: Ruru <int>, Blbj <int>, Alal <int>, Anan <int>
```

```
R> # Hellinger-transform the species dataset
R> # Suited to species abundance data. It gives low weights to species
R> # with low counts and many zeros. The transformation comprises dividing
R> # each value in a data matrix by its row sum, and taking the square root
R> # of the quotient.
R> spe.hel <- as.tibble(decostand(spe, "hellinger"))
R> spe.hel
# A tibble: 29 x 27
  Cogo Satr Phph Babl Thth Teso Chna Pato Lele Sqce Baba Albi Gogo Eslu Pefl Rham Legi Scer Cyca
<dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl>
1 0     1     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0
2 0     0.645 0.577 0.5 0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0     0
3 0     0.559 0.559 0.559 0     0     0     0     0     0     0     0     0.25 0     0     0     0     0     0     0     0     0     0     0
4 0     0.436 0.488 0.488 0     0     0     0     0.218 0     0.218 0.309 0.309 0     0     0     0     0.243 0     0     0     0     0     0
5 0     0.243 0.297 0.243 0     0     0     0.383 0.243 0     0.243 0.343 0.343 0     0     0.243 0     0     0     0     0     0     0     0
6 0     0.378 0.436 0.488 0     0     0     0.218 0.309 0     0.218 0.218 0.218 0     0     0     0     0     0     0     0     0     0     0
7 0     0.559 0.5 0.559 0     0     0     0.25 0.25 0     0     0     0     0     0     0     0     0     0     0     0     0     0     0
8 0     0     0.267 0.463 0     0     0     0     0.598 0     0     0     0     0     0     0     0     0     0     0     0     0     0     0
9 0     0.267 0.535 0.535 0     0     0     0.378 0.378 0     0.267 0     0     0     0     0     0     0     0     0     0     0     0     0
10 0.302 0.522 0.603 0.302 0.302 0     0     0     0.302 0     0     0     0     0     0     0     0     0     0     0     0     0     0
# ... with 19 more rows, and 8 more variables: Titi <dbl>, Abbr <dbl>, Icme <dbl>, Gyce <dbl>, Ruru <dbl>, Blbj <dbl>,
# Alal <dbl>, Anan <dbl>
```

```
R> # Remove empty site 8
R> env <- env[-8, ]
R> env
# A tibble: 29 x 11
  dfs alt slo flo pH har pho nit amm oxy bod
<dbl> <int> <dbl> <dbl> <dbl> <int> <dbl> <dbl> <dbl> <dbl> <dbl>
1 0.3 934 48 0.84 7.9 45 0.01 0.2 0 12.2 2.7
2 2.2 932 3 1 8 40 0.02 0.2 0.1 10.3 1.9
3 10.2 914 3.7 1.8 8.3 52 0.05 0.22 0.05 10.5 3.5
4 18.5 854 3.2 2.53 8 72 0.1 0.21 0 11 1.3
5 21.5 849 2.3 2.64 8.1 84 0.38 0.52 0.2 8 6.2
6 32.4 846 3.2 2.86 7.9 60 0.2 0.15 0 10.2 5.3
7 36.8 841 6.6 4 8.1 88 0.07 0.15 0 11.1 2.2
8 70.5 752 1.2 4.8 8 90 0.3 0.82 0.12 7.2 5.2
9 99 617 9.9 10 7.7 82 0.06 0.75 0.01 10 4.3
10 123. 483 4.1 19.9 8.1 96 0.3 1.6 0 11.5 2.7
# ... with 19 more rows
```

```
# RDA of Hellinger-transformed fish data, constrained
# by all env. vars. in the environmental dataset (sans d.f.s.)
spe.rda <- rda(spe.hel ~ ., env)
```

```
R> # Remove the 'dfs' variable from the env dataset
R> env <- decostand(env[, -1], method = "standardize")
R> env
  alt slo flo pH har
1 1.71767530 5.04893671 -1.228203403 -0.8400173 -2.38806559 -0.6297
2 1.71025775 -0.06029282 -1.219304073 -0.2734940 -2.68052294 -0.6184
3 1.64349976 0.01918408 -1.174807424 1.4260758 -1.97862529 -0.5847
4 1.42097314 -0.03758514 -1.134204232 -0.2734940 -0.80879586 -0.5285
5 1.40242926 -0.13976973 -1.128085943 0.2930293 -0.10689821 -0.2136
6 1.39130293 -0.03758514 -1.115849365 -0.8400173 -1.51069352 -0.4160
7 1.37275904 0.34844554 -1.052441640 0.2930293 0.12706768 -0.5622
8 1.04267789 -0.26466200 -1.007944992 -0.2734940 0.24405062 -0.3036
9 0.54199300 0.72312237 -0.718716775 -1.9730638 -0.22388115 -0.5734
10 0.04501688 0.06459945 -0.168070747 0.2930293 0.59499945 -0.3036
```

```
> sum(spe.rda$CA$eig)
[1] 0.1371389
```

```
> sum(spe.rda$CCA$eig)
[1] 0.3653713
```

```
> (spe.rda <- rda(spe.hel ~ ., env2)) # Observe the shortcut
Call: rda(formula = spe.hel ~ alt + slo + flo + pH + har + pho + nit + amm + oxy + bod, data = env2)
```

	Inertia	Proportion	Rank
Total	0.5025	1.0000	
Constrained	0.3654	0.7271	12
Unconstrained	0.1371	0.2729	16

Inertia is variance

Eigenvalues for constrained axes: *RDA*n eigenvalues measure amount of variance explained by the RDA

RDA1	RDA2	RDA3	RDA4	RDA5	RDA6	RDA7	RDA8	RDA9	RDA10	RDA11	RDA12
0.22808	0.05370	0.03212	0.02321	0.00870	0.00722	0.00487	0.00292	0.00214	0.00116	0.00091	0.00034

Eigenvalues for unconstrained axes: *PC*n eigenvalues measure amount of variance represented by the residual axes, *i.e.* not explained by the RDA

PC1	PC2	PC3	PC4	PC5	PC6	PC7	PC8	PC9	PC10	PC11	PC12	PC13	PC14	PC15	PC16
0.04581	0.02814	0.01528	0.01399	0.00984	0.00767	0.00420	0.00331	0.00276	0.00202	0.00175	0.00099	0.00059	0.00047	0.00021	0.00010

```
> # Proportion of variation explained by RDA1 is therefore
> spe.rda$CCA$eig[1] / (sum(spe.rda$CCA$eig) + sum(spe.rda$CA$eig))
RDA1
0.4538882
```



```
> summary(spe.rda)      # Scaling 2 (default)
```

```
Call:
rda(formula = spe.hel ~ alt + slo + flo + pH + har + pho + nit +      amm + oxy + bod, data = env2)
```

```
Partitioning of variance:
      Inertia Proportion
Total      0.5025      1.0000
Constrained 0.3654      0.7271
Unconstrained 0.1371      0.2729
```

Eigenvalues, and their contribution to the variance

Importance of components:

	RDA1	RDA2	RDA3	RDA4	RDA5	RDA6	RDA7	RDA8	RDA9	RDA10	RDA11	RDA12	PC1	PC2
Eigenvalue	0.2281	0.0537	0.03212	0.02321	0.008699	0.007218	0.004869	0.002924	0.002141	0.001160	0.000914	0.0003405	0.04581	0.02814
Proportion Explained	0.4539	0.1069	0.06392	0.04618	0.017311	0.014363	0.009689	0.005819	0.004261	0.002308	0.001819	0.0006776	0.09116	0.05601
Cumulative Proportion	0.4539	0.5607	0.62466	0.67084	0.688155	0.702518	0.712207	0.718027	0.722287	0.724596	0.726415	0.7270922	0.81825	0.87425
	PC3	PC4	PC5	PC6	PC7	PC8	PC9	PC10	PC11	PC12	PC13	PC14	PC15	
Eigenvalue	0.01528	0.01399	0.009841	0.007674	0.004201	0.003311	0.002761	0.002017	0.001752	0.0009873	0.0005919	0.0004674	0.0002128	
Proportion Explained	0.03042	0.02784	0.019583	0.015271	0.008361	0.006588	0.005494	0.004013	0.003486	0.0019647	0.0011779	0.0009301	0.0004234	
Cumulative Proportion	0.90467	0.93251	0.952090	0.967361	0.975722	0.982310	0.987804	0.991818	0.995304	0.9972687	0.9984467	0.9993767	0.9998002	
	PC16													
Eigenvalue	0.0001004													
Proportion Explained	0.0001998													
Cumulative Proportion	1.0000000													

Accumulated constrained eigenvalues

Importance of components:

	RDA1	RDA2	RDA3	RDA4	RDA5	RDA6	RDA7	RDA8	RDA9	RDA10	RDA11	RDA12		
Eigenvalue	0.2281	0.0537	0.03212	0.02321	0.008699	0.007218	0.004869	0.002924	0.002141	0.001160	0.000914	0.0003405		
Proportion Explained	0.6243	0.1470	0.08791	0.06351	0.023808	0.019755	0.013326	0.008003	0.005860	0.003175	0.002502	0.0009320		
Cumulative Proportion	0.6243	0.7712	0.85913	0.92264	0.946448	0.966202	0.979528	0.987532	0.993391	0.996566	0.999068	1.0000000		

Scaling 2 for species and site scores

* Species are scaled proportional to eigenvalues

* Sites are unscaled: weighted dispersion equal on all dimensions

* General scaling constant of scores: 1.93676

- the coordinates of the tips of the vectors representing the response variables in the biplots (or triplots); they depend on the scaling chosen

Species scores

	RDA1	RDA2	RDA3	RDA4	RDA5	RDA6
Cogo	0.13386	0.11619	-0.238205	0.018531	0.043161	-0.029728
Satr	0.64240	0.06654	0.123649	0.181606	-0.009584	0.029785
Phph	0.47477	0.07009	-0.010153	-0.115349	-0.045312	-0.030034
Babl	0.36260	0.06966	0.041311	-0.190563	-0.046944	0.006446
Thth	0.13081	0.10707	-0.239273	0.043512	0.065818	0.003468
Teso	0.06590	0.12471	-0.216916	-0.004228	0.021732	-0.004195
Chna	-0.17416	0.06774	-0.008397	-0.016402	-0.079764	0.044700
Pato	-0.12679	0.16050	-0.035692	-0.016060	-0.089638	-0.001905
Lele	-0.07961	0.04200	0.007677	-0.059155	-0.033512	-0.121451
Sqce	-0.10903	-0.17555	-0.090012	-0.168382	0.019417	0.008753
Baba	-0.18525	0.21152	-0.073101	-0.006901	-0.012999	0.040480
Albi	-0.16059	0.15514	-0.014286	-0.002463	-0.060684	0.011025
Gogo	-0.20538	0.02482	-0.007965	-0.017743	-0.049209	-0.096232
Eslu	-0.10731	0.02857	0.090028	0.012339	0.075334	-0.057091
Pefl	-0.09159	0.10509	0.070433	-0.057397	0.014063	-0.009913
Rham	-0.20905	0.16004	0.025478	0.012081	-0.011453	0.022031
Legi	-0.22798	0.11121	0.018794	-0.009470	-0.027464	0.024518
Scer	-0.16101	0.01353	0.041554	0.032367	0.054060	-0.094579
Cyca	-0.17381	0.14903	0.022239	0.009535	0.005020	-0.005397
Titi	-0.14025	0.10636	0.078279	-0.122633	0.054114	0.031262
Abbr	-0.18596	0.12223	0.053808	0.026142	0.043877	0.014596
Icme	-0.14631	0.08899	0.061818	0.034746	0.083492	0.004417
Gyce	-0.30883	0.01606	0.039351	0.029260	-0.011189	-0.052403
Ruru	-0.31982	-0.16596	-0.018193	-0.115462	0.054397	0.064760
Blbj	-0.23901	0.09089	0.051566	0.010197	0.006795	0.036516
Alal	-0.43215	-0.22643	-0.108131	0.138837	-0.083787	0.008447
Anan	-0.19440	0.14152	0.033624	0.017384	0.008122	0.017637

Site scores (weighted sums of species scores)

	RDA1	RDA2	RDA3	RDA4	RDA5	RDA6
row1	0.40149	-0.154133	0.55506	1.601005	0.193044	0.916850
row2	0.53522	-0.025131	0.43393	0.294832	-0.518997	0.458849
row3	0.49429	-0.014617	0.49415	0.169258	-0.246061	0.163409
row4	0.33451	0.001188	0.51644	-0.320793	0.089569	-0.219820
row5	0.02794	-0.194202	0.44626	-0.559104	0.855973	-1.115731
row6	0.24422	-0.130758	0.41397	-0.696109	0.182246	-0.273498
row7	0.46589	-0.126054	0.31692	-0.137640	-0.549192	-0.061715
row8	0.03660	-0.605090	-0.06989	-1.261027	0.668699	1.165074
row9	0.31380	-0.198797	0.10795	-0.635022	-0.742828	-0.990209
row10	0.48116	-0.039713	-0.37858	0.181771	0.220972	0.254669
row11	0.49162	0.014144	-0.37992	0.162929	0.223012	0.324824
row12	0.49848	0.212240	-0.67436	0.518502	0.399406	0.221762
row13	0.38203	0.229406	-0.75795	0.223281	0.515085	-0.139590
row14	0.28740	0.218549	-0.71897	-0.211161	0.175136	-0.553069
row15	0.09131	0.400116	-0.34437	-0.376171	-0.366402	-0.575447
row16	-0.05303	0.423927	-0.41000	-0.188493	-0.725181	0.151547
row17	-0.14182	0.385867	-0.36804	-0.217143	-0.643463	-0.001315
row18	-0.28203	0.275496	-0.01868	-0.371433	-0.692317	-0.062410
row19	-0.39681	0.209508	0.11549	-0.177920	-0.387207	0.048563
row20	-0.42850	0.278334	0.22003	-0.005965	-0.027098	-0.042218
row21	-0.46552	0.251911	0.22773	0.040193	0.152844	0.032242
row22	-0.28125	-1.145589	-0.50524	0.300043	-0.003393	1.157109
row23	-0.40896	-0.752956	-0.26777	0.428883	-0.647205	0.643149
row24	-0.35207	-0.770295	-0.12183	0.459218	0.079646	-1.726015
row25	-0.46923	0.094061	0.23046	0.107850	0.310149	0.132668
row26	-0.47021	0.213629	0.24872	0.084194	0.331473	0.125540
row27	-0.47266	0.234036	0.27037	0.105748	0.381225	0.093844
row28	-0.37456	0.393349	0.10404	0.202063	0.282836	0.021890
row29	-0.48931	0.321574	0.31409	0.278210	0.488026	-0.150951

Biplot scores for constraining variables

	RDA1	RDA2	RDA3	RDA4	RDA5	RDA6
alt	0.8239	-0.203027	0.46599	-0.16932	0.003151	0.10413
slo moderate	-0.3592	-0.008707	-0.21727	-0.18287	0.158087	0.50092
slo steep	0.2791	0.155999	-0.06881	0.01921	0.176534	-0.15468
slo very_steep	0.6129	-0.148526	0.45392	0.03640	-0.190954	-0.04712
flo	-0.7769	0.255048	-0.17494	0.30980	0.227429	-0.11932
pH	0.1023	0.178385	-0.30143	0.03941	0.298243	0.04856
har	-0.5722	0.044958	-0.56049	-0.14839	0.275283	-0.24521
pho	-0.4930	-0.650420	-0.19865	0.29287	0.056281	-0.39347
nit	-0.7755	-0.203755	-0.23289	0.19739	-0.078644	-0.35079
amm	-0.4744	-0.687510	-0.16641	0.28475	-0.051257	-0.33858
oxy	0.7632	0.575392	-0.16436	0.08018	-0.136461	0.13748
bod	-0.5171	-0.791730	-0.15644	0.22067	0.075935	-0.09106

Centroids for factor constraints

	RDA1	RDA2	RDA3	RDA4	RDA5	RDA6
slo low	-0.2800	0.005549	-0.09029	0.07610	-0.07882	-0.18390
slo moderate	-0.2093	-0.005073	-0.12660	-0.10656	0.09212	0.29188
slo steep	0.1965	0.109846	-0.04845	0.01353	0.12431	-0.10892
slo very_steep	0.3908	-0.094698	0.28941	0.02321	-0.12175	-0.03004

```
> # Canonical coefficients from the rda object
> coef(spe.rda)
```

	RDA1	RDA2	RDA3	RDA4	RDA5	RDA6	RDA7	RDA8	RDA9
alt	0.0004483347	7.795777e-05	0.0005188756	0.0003875664	0.001857466	-6.331179e-05	-0.001359974	0.001120126	-0.0002517503
slo moderate	-0.0123140760	-1.655649e-02	0.0160736225	-0.0278161005	0.276687997	1.310961e-01	-0.022999100	0.018861788	-0.3109131626
slo steep	0.0480170930	4.905556e-02	0.1023432587	0.1349154913	0.394849998	-1.795415e-01	0.046031368	0.123198624	0.0970066893
slo very_steep	0.0181630025	-5.708251e-02	0.2326204779	0.1005535075	0.037825142	-1.741339e-01	0.516814197	0.069166271	-0.2260049338
flo	-0.0014041126	4.456720e-03	0.0089169975	0.0164695330	0.013331778	2.705617e-03	-0.002458359	0.010400061	-0.0006270792
pH	0.0189943698	-3.167179e-02	-0.0480329994	0.1142667244	0.413629926	1.092387e-01	0.141057854	-0.436371199	-0.0219077691
har	0.0025569166	-1.952059e-03	-0.0065931826	-0.0093613897	0.005218302	-6.097975e-03	0.002166368	0.010548543	-0.0006706737
pho	0.1033308237	4.578896e-02	-0.0998238552	-0.1049627245	0.423624076	-3.693234e-01	0.037569158	-0.700230790	-0.2878854128
nit	-0.0123768964	1.045028e-01	0.0622488395	0.0772807715	0.234375571	-3.544812e-02	-0.241069952	0.128366902	-0.0685703803
amm	-0.1088564269	-4.418756e-01	0.0062834673	0.0542213439	-1.813295370	4.650524e-03	0.280777588	1.065125427	0.3119031905
oxy	0.0686523744	1.987930e-02	-0.0895382002	0.1200081482	0.031939548	3.879211e-02	-0.058254055	0.061392921	-0.0196091542
bod	0.0108347803	-2.687024e-02	-0.0254010855	0.0744748837	0.067062942	9.276105e-02	-0.019974123	0.047982742	0.0359019363

	RDA10	RDA11	RDA12
alt	0.001189022	0.0006783471	0.0009454291
slo moderate	-0.277963100	0.0402707169	-0.2974017594
slo steep	-0.445926115	0.2456799680	-0.3472509351
slo very_steep	-0.588753472	0.2475356694	-0.1844733035
flo	0.004413371	-0.0022711471	0.0064860542
pH	-0.902424704	0.0732392948	0.5765506530
har	0.003336283	0.0007591452	0.0062067627
pho	0.247283885	-0.0018071075	-0.6300023622
nit	0.113704621	0.3978086000	0.0936975559
amm	-1.224213088	-1.5939301235	0.8974262542
oxy	0.089825552	0.0623806783	0.0257096795
bod	0.065688349	0.1112209162	0.0402215529

```
> # Unadjusted R^2 retrieved from the rda object
```

```
> (R2 <- RsquareAdj(spe.rda)$r.squared)
```

```
[1] 0.7270922
```

```
> # Adjusted R^2 retrieved from the rda object
```

```
> (R2adj <- RsquareAdj(spe.rda)$adj.r.squared)
```

```
[1] 0.5224114
```

```
R> anova(spe.rda) # ... yes!
Permutation test for rda under reduced model
Permutation: free
Number of permutations: 999

Model: rda(formula = spe.hel ~ alt + slo + flo + pH + har + pho + nit + amm + oxy + bod, data = env)
      Df Variance      F Pr(>F)
Model   10  0.36889 4.9695 0.001 ***
Residual 18  0.13362
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

R>
R> # The adjusted R2 --- the variance explained by the constrained axes:
R> RsquareAdj(spe.rda)$adj.r.squared
[1] 0.5863788
```

```

R> # Variance explained by full model:
R> sum(spe.rda$CCA$eig) / spe.rda$tot.chi * 100
[1] 73.41007
R>
R> # Which axes are significant?
R> anova(spe.rda, by = "axis")
Permutation test for rda under reduced model
Forward tests for axes
Permutation: free
Number of permutations: 999

Model: rda(formula = spe.hel ~ alt + slo + flo + pH + har + pho + nit + amm + oxy + bod, data = env)
      Df Variance      F Pr(>F)
RDA1   1 0.228027 30.7183 0.001 ***
RDA2   1 0.054418  7.3309 0.001 ***
RDA3   1 0.033819  4.5559 0.049 *
RDA4   1 0.030069  4.0507 0.094 .
RDA5   1 0.007488  1.0087 1.000
RDA6   1 0.005652  0.7615 1.000
RDA7   1 0.004427  0.5963 1.000
RDA8   1 0.002816  0.3794 1.000
RDA9   1 0.001383  0.1863 1.000
RDA10  1 0.000794  0.1069 1.000
Residual 18 0.133617
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
R>
R> # Which terms are significant?
R> anova(spe.rda, by = "terms")
Permutation test for rda under reduced model
Terms added sequentially (first to last)
Permutation: free
Number of permutations: 999

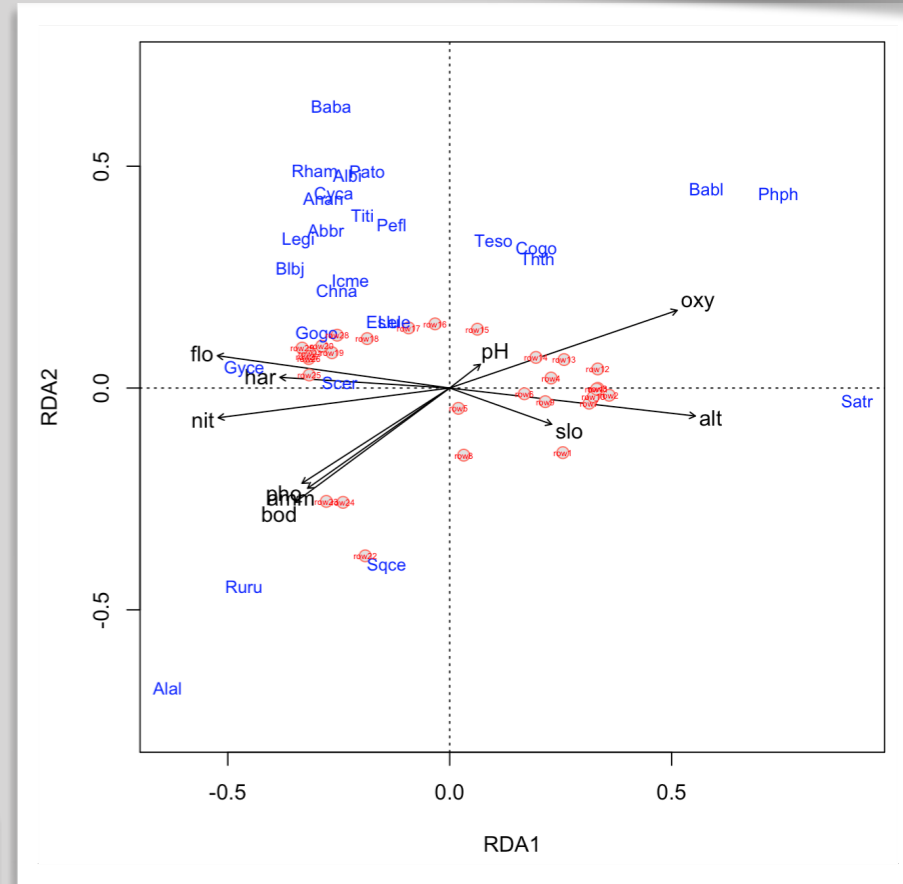
Model: rda(formula = spe.hel ~ alt + slo + flo + pH + har + pho + nit + amm + oxy + bod, data = env)
      Df Variance      F Pr(>F)
alt    1 0.164859 22.2087 0.001 ***
slo    1 0.028371  3.8219 0.004 **
flo    1 0.015431  2.0788 0.085 .
pH     1 0.011575  1.5593 0.157
har    1 0.017701  2.3846 0.066 .
pho    1 0.041819  5.6335 0.001 ***
nit    1 0.018911  2.5476 0.065 .
amm    1 0.007005  0.9437 0.426
oxy    1 0.058402  7.8675 0.001 ***
bod    1 0.004819  0.6492 0.628
Residual 18 0.133617
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```

# Another way - build from scratch:
## better control -- remember to set scaling etc. identically
plot(spe.rda, type = "n", scaling = "sites", choices = 1:2)
text(spe.rda, dis = "cn", scaling = "sites", choices = 1:2)
points(spe.rda, pch = 21, col = "salmon", bg = "grey90", cex = 1.2, scaling = "sites", choices = 1:2)
text(spe.rda, "species", col = "blue", cex = 0.8, scaling = "sites", choices = 1:2)
text(spe.rda, "sites", col = "red", cex = 0.4, scaling = "sites", choices = 1:2)

```

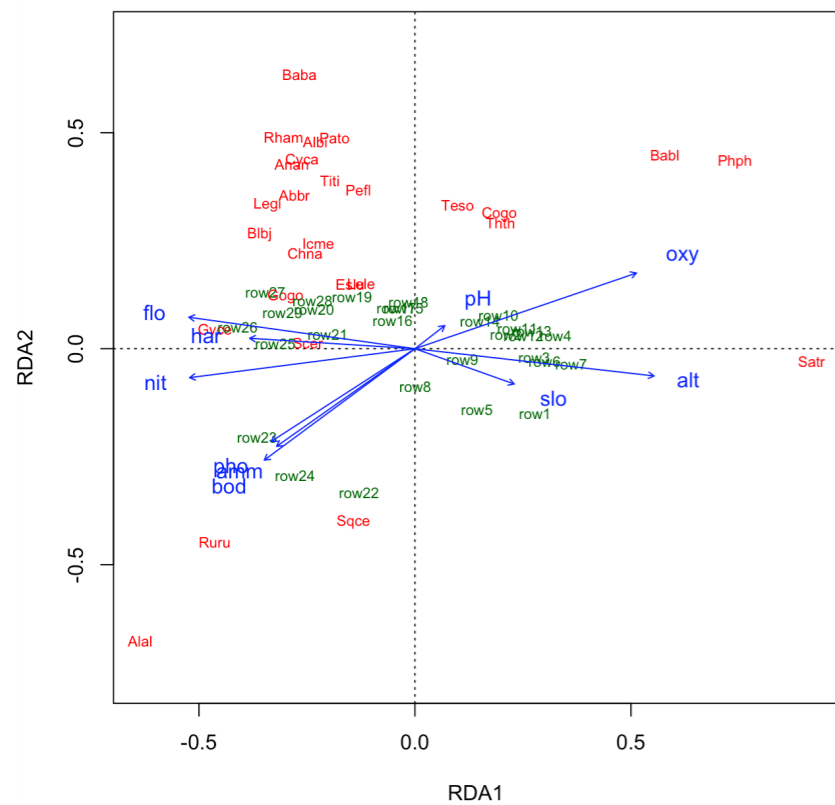


```

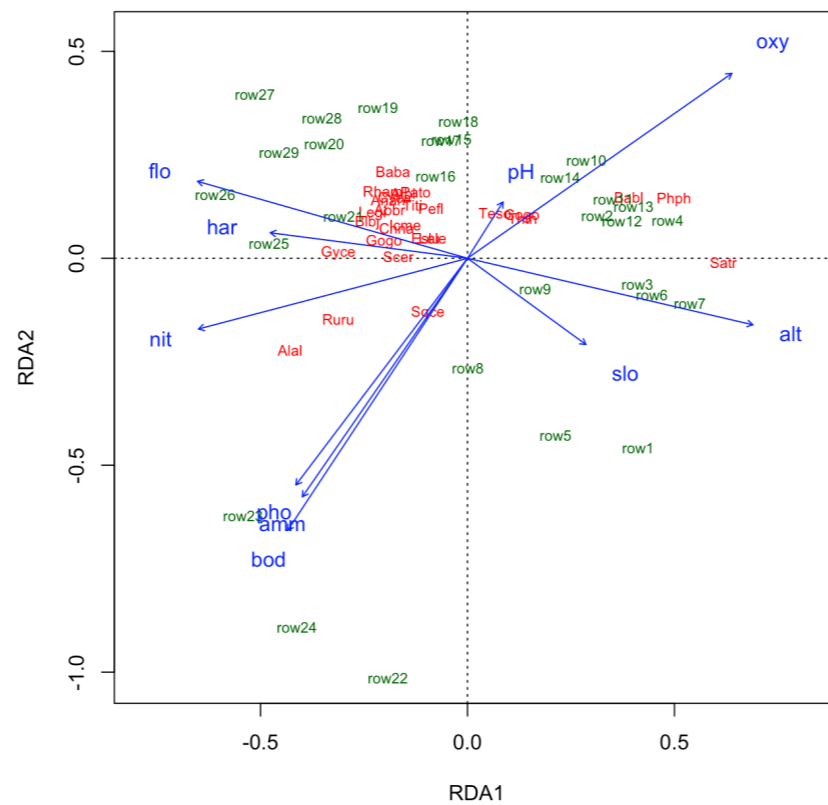
# Make some plots:
plot(spe.rda, scaling = 1, display = c("sp", "lc", "cn"),
     main = "Site scaling (scaling 1)")
plot(spe.rda, scaling = 2, display = c("sp", "lc", "cn"),
     main = "Species scaling (scaling 2)")

```

Site scaling (scaling 1)



Species scaling (scaling 2)



Canonical Correspondence Analysis (CCA)

- see <https://sites.google.com/site/mb3gustame/constrained-analyses/cca> and https://www.davidzeleny.net/anadat-r/doku.php/en:rda_cca
- CCA is the merger between CA and multiple regression, therefore also based on χ^2 -metric (dissimilarities)
- whereas CA maximises the correlation between species scores and sample scores, in CCAs the sample scores are constrained to be linear combinations of explanatory variables
- because of the 'constraint', eigenvalues in CCA will be lower than in CA
- the link to the unimodal model is clear—if a combination of environmental variables is strongly related to species composition, CCA will create a reduced axis from these variables that makes the species response curves most distinct
- second and higher axes will also maximise the dispersion (remaining inertia) of species, subject to the constraints that these higher axes are linear combinations of the explanatory variables, and that they are orthogonal (i.e. do not explain what has already been explained) to all previous axis

Canonical Correspondence Analysis (CCA)

- there are as many constrained (reduced) axes as there are explanatory variables
- the total 'explained inertia' is the sum of the eigenvalues of the constrained axes
- the remaining axes are unconstrained, and can be considered 'residual'
- the total inertia in the species data is the sum of eigenvalues of the constrained and the unconstrained axes, and is equivalent to the sum of eigenvalues, or total inertia, of CA
- thus, explained inertia, compared to total inertia, can be used as a measure of how well species composition is explained by the variables
- unfortunately, a strict measure of 'goodness of fit' for CCA is elusive, because the arch effect itself has some inertia associated with it—and it is not always clear whether this inertia belongs in the 'explained' or 'unexplained' portion

Canonical Correspondence Analysis (CCA)

- CCA benefits from the advantages of multiple regression, including:
 - it is possible that patterns result from the combination of several explanatory variables; these patterns would not be observable if explanatory variables are considered separately
 - it is possible to test hypotheses (though in CCA, hypothesis testing is based on randomisation procedures rather than distributional assumptions)
 - explanatory variables can be of many types (e.g. continuous, ratio scale, nominal) and do not need to meet distributional assumptions

Canonical Correspondence Analysis (CCA)

- Some caveats that apply to multiple regression also apply here:
 - in observational studies, not always possible to infer direct causation
 - the independent effects of highly correlated (**collinear**) variables are difficult to disentangle—however, CCA (and regression) can test the null hypothesis that such variables are completely redundant
 - it is possible to ‘overfit’ the data as the number of variables approaches the number of samples (instead of $r^2 \sim 1$, the explained inertia will equal the total inertia and the CCA solution equals the CA solution), so the solution is no longer ‘constrained’ by the variables
 - noise in explanatory variables will have an effect on the predicted values
 - the interpretability of the results is directly dependent on the choice and quality of the explanatory variables; and on the researcher’s expert knowledge
 - although both multiple regression and CCA find the best linear combination of explanatory variables, they are not guaranteed to find the true underlying gradient (which may be related to unmeasured or unmeasurable factors), nor are they guaranteed to explain a large portion of variation in the data—again expert knowledge necessary

Canonical Correspondence Analysis (CCA)

- one of the biggest advantages of CCA lies in the intuitive nature of its ordination diagram, or triplot (you've seen this already when we did the 'envfit'); it is called a triplot because it simultaneously displays three pieces of information: samples as points, species as points, and environmental variables as arrows (or points)
- CCA triplots can get very crowded, and solutions for this include:
 - separate the parts of the triplot into biplots or scatterplots (e.g. plotting the arrows in a different panel of the same figure)
 - rescaling the arrows so that the species and sample scores are more spread out
 - only plotting the most abundant species (but remember the rare ones as knowledge of these is important in certain studies)
 - plotting only significant constraints as per PERMANOVA or some other means
- please read "Environmental variables in CCA" at <http://ordination.okstate.edu>

Canonical Correspondence Analysis (CCA)

- in **vegan**'s `cca()`, a chi-square transformed data matrix is subjected to weighted linear regression on constraining variables, and the fitted values are submitted to correspondence analysis performed via singular value decomposition (svd)

db-RDA

- in **vegan**'s `capscale()`, any dissimilarity matrix from `vegdist()` is subjected to weighted linear regression on constraining variables, and the fitted values are submitted to a PCoA
- the general benefits of a PCoA also apply here
- (dis)similarity matrices calculated from quantitative, semi-quantitative, qualitative, and mixed variables can be handled
- access to a wide array of dissimilarities make the method suitable to many kinds of ecological data
- since we apply species information is lost during the calculation of the dissimilarity matrix, if the original matrix of species composition matrix is available, the species scores can be added into the final ordination diagram as weighted means of site score in which they occur or as vectors fitted onto the ordination space

PERMANOVA vs. ANOSIM

- “Overall, ANOSIM and the Mantel test were very sensitive to heterogeneity in dispersions, with ANOSIM generally being more sensitive than the Mantel test. In contrast, PERMANOVA and Pillai’s trace were largely unaffected by heterogeneity for balanced designs. [...]. PERMANOVA was also unaffected by differences in correlation structure. [...] PERMANOVA was generally, but not always, more powerful than the others to detect changes in community structure...”